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and Insulin

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## Introduction:

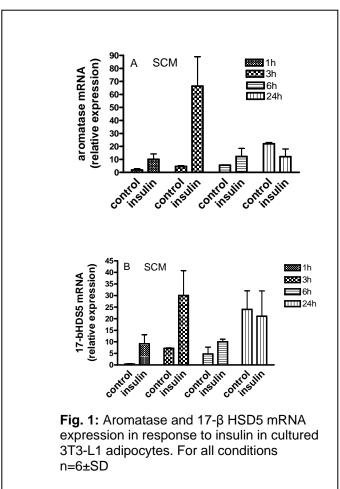
Clinical studies suggest and obesity increases the risk for breast cancer and there is convincing evidence that post-menopausal breast cancer risk is highly correlated with serum estrogen levels. One potential link between obesity and breast cancer risk is increased estrogen production by the adipose tissue itself. The adipose tissue produces the enzyme aromatase which catalyses the biosynthesis of estrogen from androgen and also 17- $\beta$ -hydroxysteroid dehydrogenase (17- $\beta$ HSD) important for the conversion of estrone to estradiol. In spite of this the mechanisms regulating the adipose expression of aromatase and 17- $\beta$ HSD is however currently unknown. Identifying the mediators in obesity that regulate aromatase and 17- $\beta$ HSD synthesis in adipose tissues and in adipocytes may provide a unique therapeutic preventive strategy to reduce systemic estrogen levels and thereby reduce post-menopausal breast cancer risk associated with obesity.

### **BODY:**

# Task 1 : Perform in vitro studies on the regulation of Aromatase and 17- $\beta$ HSD (types 4, 5) synthesis in murine and human adipocytes in response to insulin and leptin

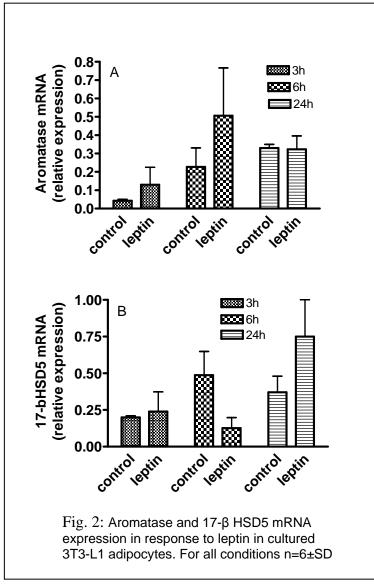
In order to determine the regulation of Aromatase and 17-βHSD in adipocytes we initially standardized an in vitro murine adipocyte cell culture system, In this model, 3T3, L1 pre adipocytes were grown and differentiated into adipocytes after a brief exposure of confluent pre-adipocytes to insulin and dexamathasone. This treatment triggered the differentiation of pre adipocytes to fully differentiated

lipid filled adipocytes over the course of 2-3 weeks. Fully differentiated 3T3-L1 adipocytes were treated with insulin (100nM), and cells harvested at various times after treatment for total RNA extraction. Aromatase and 17-βHSD5 mRNA expression was determined by real time RT-PCR. Treatment of fully differentiated 3T3-L1 adipocytes with 100nM insulin in serum containing media (SCM) significantly induced both Aromatase (Fig 1 A) and 17-βHSD5 mRNA (Fig 1 B) expression in these cells. Aromatase mRNA expression was induced significantly as early as 1 hour after insulin treatment and this expression continued to increase by 3 h after insulin treatment. Similar kinetics of induction of mRNA was also observed after insulin treatment for the expression of 17-βHSD5 mRNA (Fig. 1B). 17-βHSD5 mRNA expression was dramatically induced at 1 and 3 h after insulin treatment. These studies suggest that hyperinsulinemia associated with obesity may contribute to the increased expression of Aromatase and 17-BHSD5 mRNA from adipocytes.



In a different set of experiments fully differentiated 3T3-L1 adipocytes were treated with leptin (100nM) and the kinetics of both Aromatase and 17- $\beta$ HSD5 mRNA regulation determined at 3, 6, and 24 hrs after leptin treatment. Aromatase mRNA expression was significantly induced in adipocytes at 3 and 6 hrs after leptin treatment (Fig. 2A). In contrast to Aromatase mRNA expression, the gene expression of  $\beta$ HSD5 was reduced 3 hrs after leptin treatment but was significantly increased after exposure to leptin for 24 hrs. These data suggest that long term chronic exposure to leptin may increase the expression of  $\beta$ HSD5 from adipocytes.

Since human obesity is associated with elevated levels of leptin, our data suggest that hyperleptinemia associated with obesity may contribute not only to Aromatase gene expression but also to increased levels of  $\beta$ HSD5 in adipocytes. These results thus support our primary hypothesis that the hyperleptinemia and hyperinsulinemia associated with obesity may induce the expression of Aromatase and  $\beta$ HSD5 from the adipose tissue, specifically from adipocytes. Studies to determine whether the increase in Aromatase and  $\beta$ HSD5 gene expression in response to insulin and leptin actually leads to increased estrogen secretion into the conditioned media are ongoing.

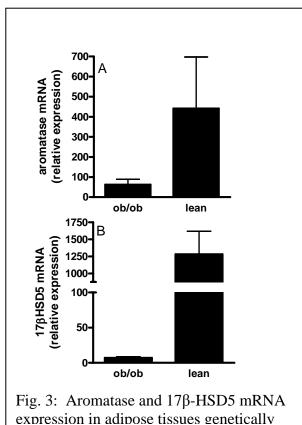


obesity.

Since obesity is usually associated with insulin resistance we had also proposed to determine the induction of Aromatase and βHSD5 mRNA expression in response to insulin and leptin in insulin resistant adipocytes. 3T3-L1 adipocytes were treated with low doses (2ng/ml) of tumor necrosis factor  $\alpha$  consecutively for 3 days. Metabolic insulin resistance was determined by measuring insulin mediated glucose uptake. Glucose uptake was reduced by 50-70% in adipocytes treated with TNF-α compared to untreated cells. suggesting that glucose uptake was blunted in these cells. Thus, we have been able to standardize conditions in our cell culture system to mimic metabolic insulin resistance. These "insulin-resistant" adipocytes are currently being used to determine insulin and leptin mediated regulation of Aromatase and βHSD5 expression by these cells. Preliminary studies indicate that these "metabolically insulin resistant" adipocytes remain sensitive to insulin in terms of aromatase and 17β HSD5 induction. These results suggests that the signaling pathways by which insulin induces aromatase and 17βHSD5 expression do not become "insulin resistant" but continue to respond to the hyperinsulinemia associated with

Task 2: Perform in vivo studies on the regulation of Aromatase and 17-beta HSD (types 4, 5) expression in response to leptin and/or insulin using lean, diet-induced obese and genetically obese mice.

While our hypothesis was that increased insulin and leptin levels associated with obesity may drive the up regulation of both aromatase and 17-βHSD expression in adipose tissues, we had not in fact directly compared the expression of these genes in adipose tissues of lean and obese mice. We used both the genetically obese ob/ob mice, and a diet induced model of obesity in normal C57BL/6J mice to determine aromatase and 17-BHSD expression in adipose tissues. In the model of diet induced obesity lean C57BL/6J mice were placed either on a high fat diet (HFD; 60% kcal from fat) or a low fat diet (10% kcal from fat) for 16 weeks. On the HFD the mice became obese, insulin resistant and diabetic. The genetically obese ob/ob mice is a severe model of obesity and these mice also lacks the satiety hormone leptin. The diet induced model of obesity is a milder and a more physiologically relevant form of obesity which better mimics the human condition. Moreover, in contrast to the genetic model, the HFD-induced obese mice also show increased levels of leptin which is similar to human obesity. In the genetically obese (ob/ob) mice, the expression levels of both aromatase and 17β-HSD were in fact reduced in ob/ob mice when compared to its lean counterparts (Fig. 3). However, when we determined the levels of these genes in the adipose tissues of diet-induced obese mice there was a significant increase in both the aromatase and 17β-HSD expression in mice fed the HFD compared to those on the LFD (Fig. 4). Based on the results we obtained in these two models of obesity, we hypothesized that the decrease in the expression of aromatase and 17β-HSD5 in adipose tissues of the



600 relative expression aromatase mRNA 400· 300-10 **HFD** LFD 600-В (relative expression) 17B HSD5 mRNA 400 300 200 100 **HFD LFD** 

Fig. 4: Aromatase and 17β-HSD5 mRNA expression in adipose tissues of mice fed either a high fat diet (HFD) or a low fat diet (LFD) for 16 weeks. N=6±SD.

ob/ob mice is probably due to the lack of leptin, and, that increased leptin associated with obesity such as in the diet induced model of obesity may actually lead to elevated levels of adipose aromatase and

17-βHSD5 expression. We tested this hypothesis by injecting ob/ob mice with exogenous leptin and measured the expression of aromatase and 17β-HSD5 in the adipose tissue 3 hours after treatment. For these experiments, groups (n=6) of ob/ob mice were injected with leptin (10µg/mouse). 3 hours later, mice were sacrificed and blood and adipose tissues were harvested. Total RNA was extracted from adipose tissues and the expression of Aromatase and 17-BHSD4 mRNA expression determined by real time RT-PCR. As indicated in Figure 5, leptin treatment of the leptin deficient ob/ob mice led to a dramatic increase in the expression of both aromatase and 17β-HSD5 mRNA levels in the adipose tissues. Leptin treatment of lean mice similarly induced the adipose expression of aromatase 7-BHSD4 mRNA (Fig. 6, 7).

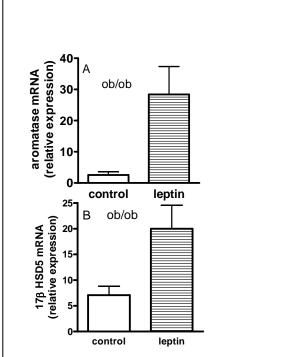
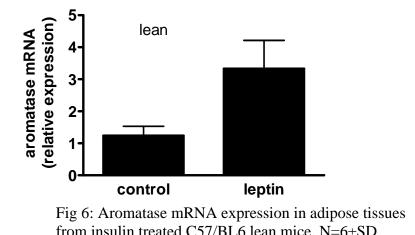


Fig. 5: Aromatase and 17β-HSD5 mRNA expression in adipose tissues of genetically obese ob/ob treated with leptin. N=6±SD.

These studies support our hypothesis that hyperleptinemia associated with obesity leads to the up regulation of aromatase and 17β-HSD5 mRNA in adipose tissues.

We next performed experiments to



from insulin treated C57/BL6 lean mice. N=6±SD

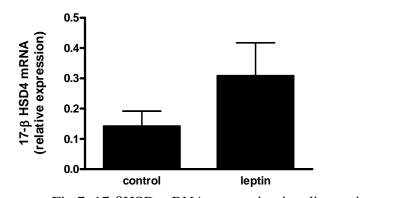


Fig 7: 17-βHSD mRNA expression in adipose tissues from insulin treated C57/BL6 lean mice. N=6±SD

determine whether insulin induces the expression of Aromatase and 17-βHSD expression in adipose tissues in vivo. Groups (n=6) of C57BL/6J lean mice were injected with insulin (humulin, 5 IU), and 3 hours later, mice were sacrificed and blood and adipose tissues harvested. Total RNA was extracted from adipose tissues and the expression of Aromatase and 17-βHSD mRNA expression determined by real time RT-PCR. As shown in Fig 8, insulin treatment also induced a dramatic and significant expression of aromatase mRNA in adipose tissue of lean mice. Our preliminary studies had previously shown that Aromatase gene expression is also induced by insulin in the obese, ob/ob mice that are insulin resistant. Studies are ongoing to confirm and extend the results relating to the insulin induction of aromatase gene expression in adipose tissues from insulin resistant genetic and diet-induced obese mice.

We next determined the expression of 17-βHSD mRNA in adipose tissues of insulin treated C57BL/6J lean mice. Interestingly, the expression of 17-βHSD5 mRNA levels was very low in the adipose tissues of these mice and hence we were unable to make any meaningful conclusions in relation to its

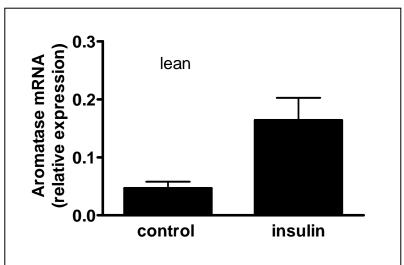


Fig. 8: Aromatase mRNA expression in adipose tissues From insulin treated C57/BL6 lean mice. N=6±SD

genetic and diet-induced obese mice.

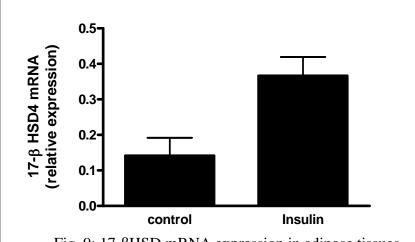


Fig. 9: 17- $\beta$ HSD mRNA expression in adipose tissues from insulin treated C57/BL6 lean mice. N=6 $\pm$ SD

expression. However, we show that 17-βHSD4 mRNA was expressed in adipose tissues of control untreated C57BL.6J mice, and its expression was significantly and dramatically induced 3 hr after insulin treatment (Fig. 9). These data suggest that the hyperinsulinemia associated with insulin resistance and obesity may drive the expression of both Aromatase and 17-βHSD mRNA and thereby contribute to increased estrogen secretion from adipose tissues in obesity. Studies are ongoing to confirm and extend the results relating to the insulin induction of 17-βHSD gene expression in adipose tissues from insulin resistant

Together, our in vivo data suggest that increased elevated levels of insulin and leptin, associated with obesity may increase breast cancer risk by inducing the adipose expression of both Aromatase and 17-βHSD. While Aromatase catalyses the biosynthesis of estrogens from androgens, 17βHSDs are important for the conversion of estrone to estradiol (1). Thus the increased expression of both of these enzymes is important for the production of biologically active estrogen. Are data suggest that insulin and leptin are important in this process.

# **Key research Accomplishments:**

- Aromatase and 17-β HSD5 mRNA expression is increased in the adipose tissues of genetically obese ob/ob mice that are leptin deficient compared to its lean counterpart.
- Aromatase and 17-β HSD5 mRNA levels are increased in the adipose tissues of mice fed a high fat diet compared to those on a low fat diet.
- Aromatase and 17-βHSD4 gene expression is increased in vivo in adipose tissues of ob/ob and C57BL/6J lean mice treated with **leptin**
- Aromatase and 17-β HSD5 mRNA expression is also increased in response to **leptin** in cultured 3T3-L1 adipocytes.
- Aromatase and 17-βHSD4 gene expression is increased in vivo in adipose tissues of C57BL/6J lean mice treated with **insulin**
- Aromatase and 17-β HSD5 mRNA expression is also increased in response to insulin in cultured 3T3-L1 adipocytes.

The results of our studies have identified two key mediators in obesity (insulin and leptin) that regulates aromatase and 17- $\beta$ HSD synthesis in the adipose tissue and in adipocytes. The results from this study may provide a unique therapeutic prevention strategy to reduce systemic estrogen levels and thereby reduce postmenopausal breast cancer risk associated with obesity.

**Reportable outcomes:** Manuscript in preparation.

### **Conclusion:**

In conclusion, our studies support the hypothesis that hyperleptinemia and hyperinsulinemia associated with obesity can induce not only aromatase but also 17-β HSD synthesis from the adipose tissue which may lead to an increase secretion of estrogen from an expanded adipose tissue in obesity. These studies provide a potential molecular link by which obesity leads to increased risk for breast cancer. Targeting the adipose production of enzymes leading to elevated estrogen production may provide a feasible therapeutic option to reduce breast cancer risk.

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